Chromatin structure abnormalities of human spermatozoa. Implications for infertility

Anomalies de structure de la chromatine des spermatozoïdes humains Implications pour l'infertilité

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Landmarks of in vitro fertilisation technology (IVF)

First attempts at in vitro fertilisation in 1878 mammalian eggs 1880 First successful embryo culture 1930 First successful ivf of mammalian eggs resulting in a live birth 1935 First successfully fertilised human eggs in vitro 1978 **Birth of Louise Brown** 1990 **Preimplantation diagnosis** 1992 Intracytoplasmatic sperm injection developed in humans

Landmarks II

Partial zona dissection (PZD)
Sub Zonal Insemination (SUZI)

Intracytoplasmatic sperm injection (ICSI)

 Rounded spermatid nucleus injection (ROSNI)

Testicular sperm aspiration (TESA) or biopsy (TESE)

Microsurgical Epididymal Sperm Aspiration (MESA) Skipping evolutionary barriers? Is it going to affect the future?

> "Bad genes tend to end up in bodies that die young or without reproducing (R.Dawkins,1995)

"What form of male infertility are we left to cure? (Silber, 1995)

Risks for off-spring and assisted reproductive technology. Questions are now being asked... What do we know?



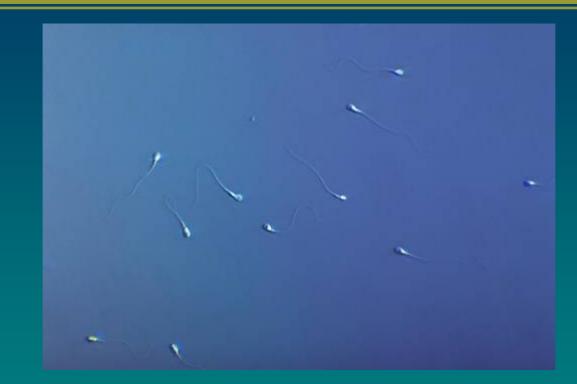
Seeds of dout Powell K: Nature, 17 April 2003

Twice as high a risk of a major birth defect as naturally conceived infants? Hansen et al. NEJM: 346, 725. 2002 The course of pregnancy or the outcome after ICSI are not affected by origin or number of sperm in the ejaculate Ludwig M et al. Hum. Reprod: 18, 351. 2003 ART is associated with a human overgrowth syndrome (Beckwith-Wiedemann) DeBaun MR et al. Am. J. Hum. Genet. 72, 156. 2003

Effects of sperm damage

- **Fertilisation**
- Cell division
- Embryo development
- Pregnancy outcome
- Congenital anomalies in babies
- Future health issues

Sperm characteristics



Human sperm quality is usually defined to by standard WHO semen analysis parameters: number, motility morphology WHO laboratory manual for the examination of human semen and sperm –cervical mucus interaction. 1999 Cambridge University Press

Morphology I









Amorphous









Pyriform

Macrocephalic



Microcephalic



Round





Morphology II



Mid piece anomalies





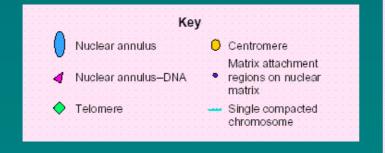


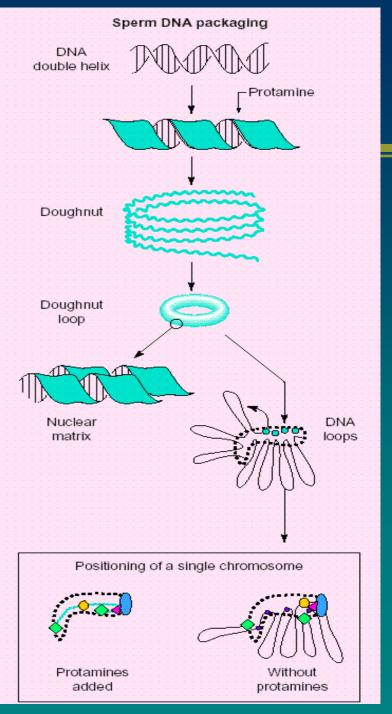
Tail anomalies



Chromatine structure in spermatozoa

Reviews of Reproduction (1999) 4, 31–37
Origin of DNA damage in ejaculated human spermatozoa
Denny Sakkas, Ewa Mariethoz, Giancarlo Manicardi, Davide Bizzaro, Patrizia G. Bianchi and Umberto Bianchi





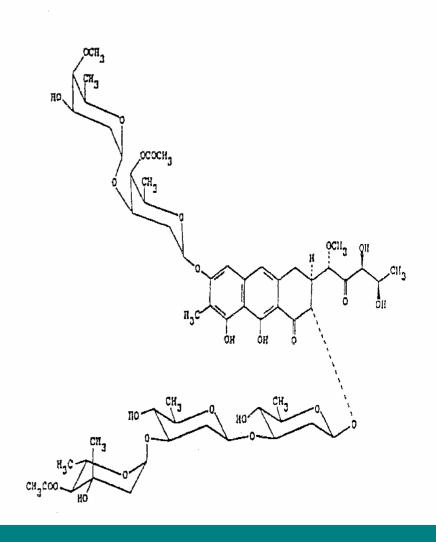
Aim of the research project

- Study chromatin structure and DNA organisation in spermatozoa
- Relate nuclear anomalies to the classical descriptive parametres of sperm quality
- Relate these anomalies to success rates of in vitro fertilisation and intracytoplasmatic sperm injection
- Isolate spermatozoa of a better nuclear consistency to increase the likehood of a normal offspring of infertile couples

Chromomycin A₃ (CMA₃)

- Fluorochrome specific for GC sequences
- Reciprocal inhibitory competitive process between protamines and CMA₃
- Chromomycin (CMA₃) is a useful tool to for assessing the packaging quality of the chromatin in spermatozoa and allows indirect visualisation of protamin deficiency

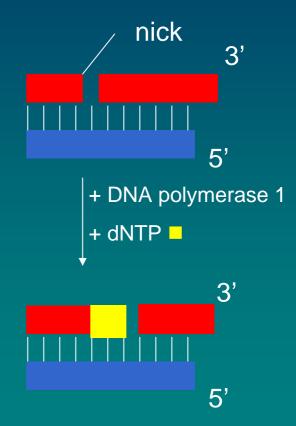
Bianchi et al. Biol. Reprod. 49, 1993



Endogenous nick translation

No endonuclease treatment prior to nick translation Detection system: streptoavidin fluorescein isothiocynate(FICT) or digoxigenin-11-dUTP

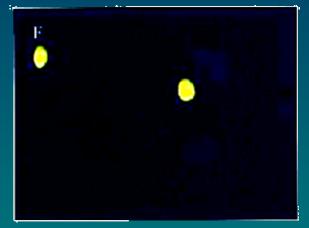
Manicardi et al. Biol. Reprod. 52, 1995



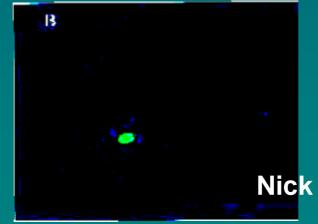
Techniques for the evaluation of sperm chromatin damage I



Phase contrast

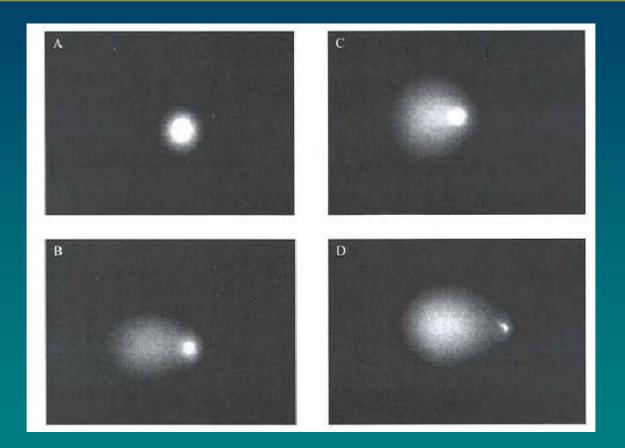






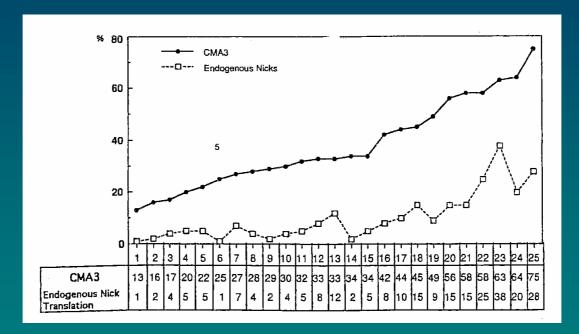
Nick translation

Different patterns of comets obtained from single spermatozoa.



The morphology and dimension of the tails are related to the amount of DNA fragments.

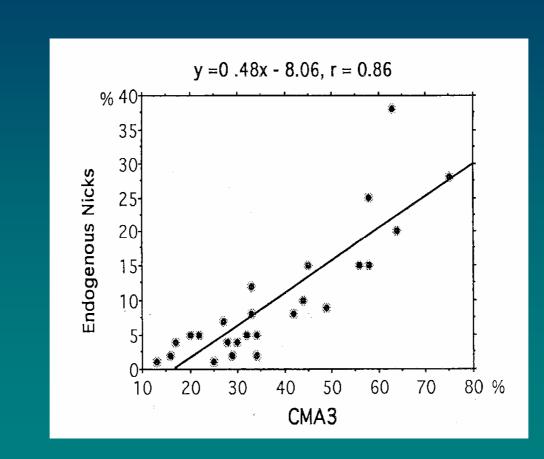
Percentage of human spermatozoa positive after treatment with CMA₃ and in situ nick translation



Values represent assessment of 500 sperm on each slide.

Manicardi et al. Biol. Reprod. 49, 1995

Correlation between CMA₃ fluorescence and presence of endogenous nicks in human spermatozoa collected from 25 patients



Manicardi et al. Biol. Reprod. 49, 1995

Relationship between CMA₃ positivity, presence of endogenous nicks and sperm morphology

In individuals presenting less than 20% of morphologically normal spermatozoa both CMA₃ and endogenous nick-translation positivity were high

Bianchi et al. J Assist. Reprod.Genet. 13, 1996

Evidence of hidden anomalies in normal spermatozoa

Infertile males undergoing sperm microinjection treatment do also show fluorescence in spermatozoa judged as morphologically normal thus suggesting the presence of hidden structural chromatin defects even in those gametes that have more chance to be selected for the procedure

Bianchi et al. Mol. Hum. Reprod. 2, 1996

Mean fertilisation rates in relation to CMA₃ positivity of morphologically normal spermatozoa

| Μ | % OF ORPHOLOGICALLY NORMAL SPERM FLUORESCING | No of cases (a) | Mean fertilisation rate (b) |
|---|--|-----------------|--------------------------------|
| | 0-50 | 10 | 56.4 <u>+</u> 25.2 |
| | 50-70 | 7 | 53.6 <u>+</u> 30.2 |
| | >70 | 10 | 24 <u>+</u> 15.9 (c) |

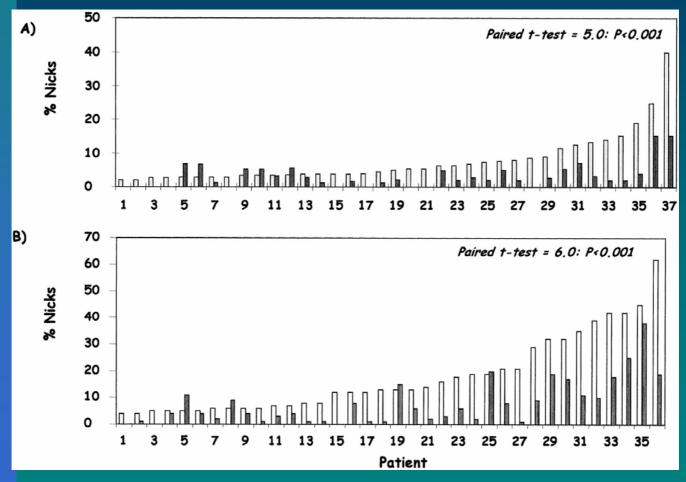
Bianchi et al. Mol. Hum. Reprod. 2, 1996

Percentage of CMA₃ positive spermatozoa after isolation using swim-up, Percoll and PureSperm techniques

| N° of samples | Fraction of sperm preparation | | | Paired t-test (wash vs s-u or 90%) |
|---------------|-------------------------------------|----------------------------------|----------------------------------|---------------------------------------|
| | Swim-up | | | |
| 31 | Wash | Sediment | Swim-up | |
| | 21.5 <u>+</u> 9.5 | 19.6 <u>+</u> 9.7 | 22.0 <u>+</u> 9.5 | P=0.6 |
| | Percoll | | | |
| 39 | Wash | 45% | 90% | |
| | 29.9 <u>+</u> 15.5 | 30.72 <u>+</u> 18.3 | 18.1 <u>+</u> 12.1 | P<0.001 |
| | Puresperm | | | |
| 45 | Wash 33.9 <u>+</u> 21.2 | 45% 25.0 <u>+</u> 19.8 | 90% 12.4 <u>+</u> 12.6 | P<0.001 |

Sakkas et al. Hum. Reprod: 15, 2000

Percentage of spermatozoa exibiting endogenous Dna nicks after preparation with Percoll (A) and Puresperm (B)



Sakkas et al. Hum. Reprod. 15, 2000

Cervical mucus characteristics I

- A heterogenous secretion containing 90% water
- **Reological properties**
 - Consistency
 - Spinnbarkeit
 - Ferning

A hydrogel comprising a high viscosity component and low viscosity

Cervical mucus characteristics II

High viscosity component:

Macromolecular network of mucins

A fibrillar system of subunits with a peptide core and oligosaccharide side chains

Low viscosity component:

– Electrolytes, organic coumpounds, soluble proteins

Cervical mucus properties

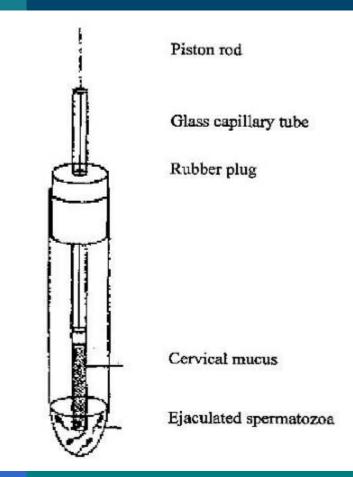
- Receptivity to sperm
- Protection of sperm from the hostile environment of the vagina
- Supplementation of energy requirements of sperm
- Short term sperm reservoir
- Initiation of sperm capacitation
- Filtering effect

– Mortimer et al., 1982; Katz et al.,1990; Eggert-Krause et al., 1995

WHO laboratory manual for the examination of human semen and sperm-cervical mucus interaction , 1992

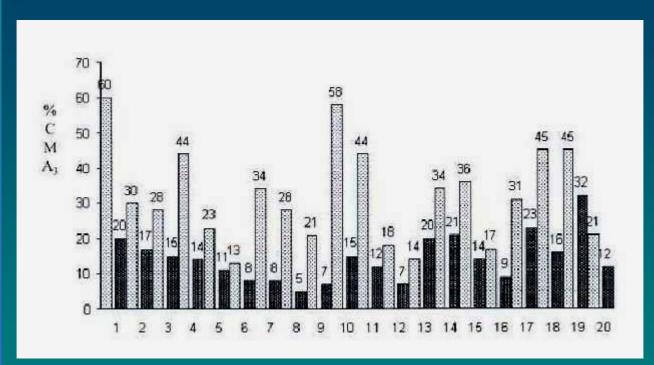
Patient selection for the cervical mucus study

- Patients were deliberately chosen on a random basis among the men consulting for couple infertility at the Sterility Clinic of the University Hospital of Geneva.
- The aim was to evaluate selection power of cervical mucus on sperm independently from classical sperm parameters.
- The controls were intra-individual as the ejaculate of each individual was separated in two and analysed.

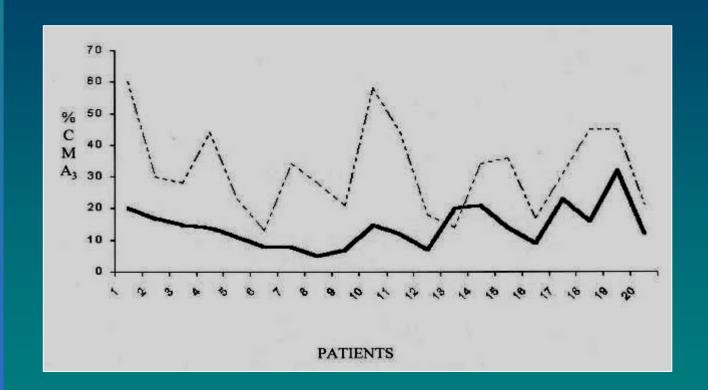


T6 medium was used at the end of the capilary tube to recover the sperm crossing successfully the mucus

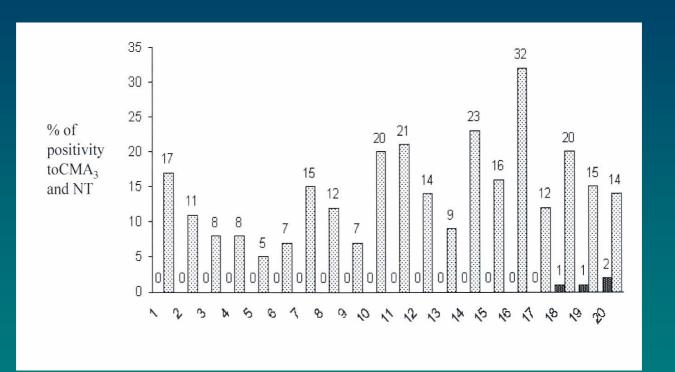
Recovered sperm was then fixed in Carnoy, spead on slides and treated according to the chromomycin and the nick translation technique protocol



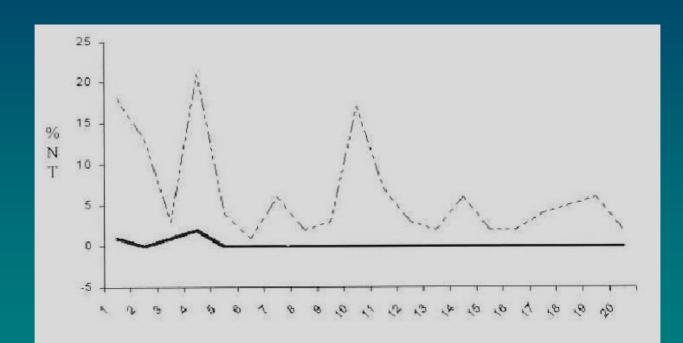
Percentage of positivity to CMA₃ in raw spermatozoa (white) and in spermatozoa selected by cervical mucus (black). T-test = 5.2256, p<0.05 = 2.024, p>0.001 = 3.566



Comparison between percentage of positive to CMA₃ of raw (-.-) and selected spermatozoa (-)



There is no correlation between percentage of positivity to in situ nick translation (black columns) and CMA₃ (white columns) estimated in spermatozoa selected by cervical mucus - r = 0.08817 - p = 0.717



Comparison between percentage of positivity to in situ nick translation of raw (-.-) and selected spermatozoa (-)

Conclusions

Number, motility and morphology only partially define the genetic quality of human spermatozoa

Evaluation of sperm quality also partially benefits from the use of techniques such as nick translation and CMA₃

Cervical mucus, in vitro, seems to be also capable of selecting structurally normal spermatozoa

Identifing sperm with physical and/or chemical integrity may allow treatment of the most severe cases of male infertility with less risks for progeny

Thank you

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